# East African pigs have a complex Indian, Far Eastern and Western ancestry

# A. Noce\*, M. Amills\*, A. Manunza\*, V. Muwanika<sup>†</sup>, D. Muhangi<sup>‡</sup>, T. Aliro<sup>§</sup>, J. Mayega<sup>†</sup>, R. Ademun<sup>¶</sup>, A. Sànchez\*, S. Egbhalsaied\*\*, A. Mercadé<sup>††</sup> and C. Masembe<sup>‡‡</sup>

\*Department of Animal Genetics, Center for Research in Agricultural Genomics (CSIC-IRTA-UAB-UB), Campus de la Universitat Autònoma de Barcelona, Bellaterra 08193, Spain. <sup>†</sup>Molecular Genetics Laboratory, Department of Environmental Management, College of Agricultural and Environmental Sciences, Makerere University, P.O.Box 7026, Kampala, Uganda. <sup>‡</sup>Department of Wildlife and Aquatic Resources, School of Veterinary Medicine and Animal Resources, College of Veterinary Medicine, Animal Resources and Biosecurity (COVAB), Makerere University, P. O. Box 7062, Kampala, Uganda. <sup>§</sup>Directorate of Production and Marketing, Gulu District Local Government, P. O. Box 2, Gulu, Uganda. <sup>¶</sup>Ministry of Agriculture Animal Industry and Fisheries, National Animal Disease Diagnostics and Epidemiology Centre, P. O. Box 513, Entebbe, Uganda. \*\*Young Researchers and Elite Club, Isfahan (Khorasgan) branch, Islamic Azad University, Isfahan, Iran. <sup>††</sup>Departament de Ciencia Animal i dels Aliments, Universitat Autònoma de Barcelona, Bellaterra, 08193, Spain. <sup>‡‡</sup>Department of Biological Sciences, School of BioSciences, Makerere University, Box 7062, Kampala, Uganda.

# Summary

In this study, we have characterized the mitochondrial diversity of 81 swine from Uganda. Median-joining network analysis of D-loop sequences from these individuals and others characterized in previous studies allowed us to determine that Ugandan pigs cluster with populations from the West (Europe/North Africa), Far East and India. In addition, partial sequencing of the Y-chromosome UTY locus in 18 Ugandan domestic pigs revealed the segregation of a single HY1 lineage that has a cosmopolitan distribution. A Western and Far Eastern ancestry for East African pigs had been already reported, but this is the first study demonstrating an additional contribution from the Indian porcine gene pool. This result is consistent with the high frequency of zebuine alleles in cattle from East Africa. The geographic coordinates of East Africa, at the crossroads of many trading routes that, through the ages, linked Europe, Africa and Asia, might explain the rich and complex genetic heritage of livestock native to this area.

Keywords genetic diversity, indian ancestry, introgression, ugandan pigs

The history of pig domestication and breeding in Africa is mostly unknown because of a lack of sufficient zooarcheological and genetic data (Blench 2000; Amills *et al.* 2013). It has been generally assumed that pigs were not raised in East Africa until the arrival of the Portuguese sailors (Blench 2000). However, the analysis of Y-chromosome variation of pigs from Zimbabwe and Kenya revealed the presence of one haplotype that segregates at high frequencies in Asian pigs, whereas it is almost absent in European standard commercial breeds (Ramírez *et al.* 2009). This result points to the entry of Asian pigs into East Africa, maybe as a consequence of the ancient Indian Ocean trade between this region and China or because of a more recent

Address for correspondence

Accepted for publication 25 March 2015

introduction coming from the Portuguese colony of Macau (Ramírez *et al.* 2009). The aim of the current study was to characterize the genetic diversity of pigs from Uganda and make general inferences about the ancestry of East African swine.

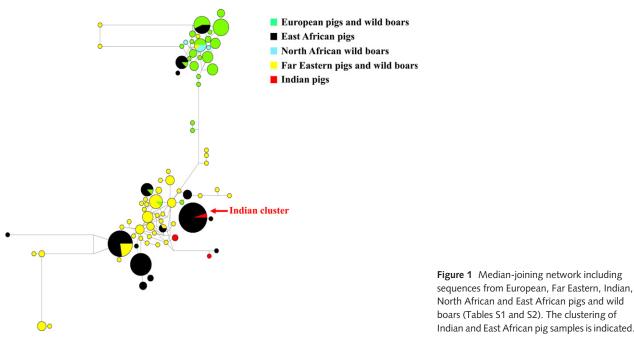
Indigenous Ugandan pigs are black, and they are characterized by their hardiness, being bred under small-holder farming and scavenging conditions. Whole blood samples were obtained from 81 local pigs distributed in 13 different locations (Table S1). Total genomic DNA was extracted with the DNeasy Blood & Tissue Kit (Qiagen). A set of primers (forward 5'-CAACCAAAACAAGCATTCCA-3', reverse 5'-GATTGTGGGCGTATGCTTAAA-3') was used to amplify 1.2 kb (GenBank reference sequence AJ002189) corresponding to the mitochondrial control region. A second pair of primers (forward 5'-CAACCAAAACAAGCAATCAAGCATTCCA-3', reverse 5'-CCCGTAACCATTGACTGAAT-3'), yielding a 0.69-kb product, was used in the few cases when the aforementioned 1.2 kb-PCR did not work. Polymerase chain reactions were carried out in a 15-µl

M. Amills, Department of Animal Genetics, Center for Research in Agricultural Genomics (CSIC-IRTA-UAB-UB), Campus de la Universitat Autònoma de Barcelona, Bellaterra, 08193 Spain. E-mail: marcel.amills@uab.cat

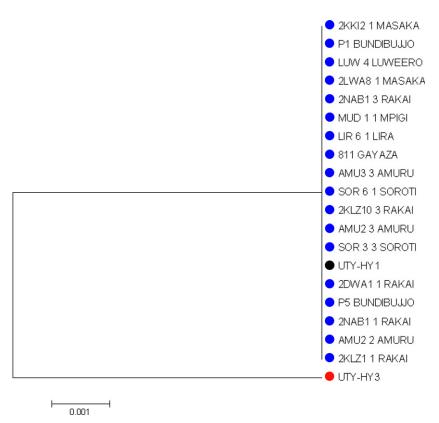
final reaction mixture containing PCR buffer, 2.5 mm of MgCl<sub>2</sub>, 0.25 mM of dNTPs, 0.3 µM of each primer and 0.75 U of Taq Gold DNA polymerase (Applied Biosystems). The thermocycling profile consisted of one cycle at 95 °C for 10 min, followed by 35 cycles of 95 °C for 1 min, 62 °C for 1 min, 72 °C for 1 min and a final extension step at 72 °C for 7 min. Amplified products were purified with the ExoSAP-IT PCR Cleanup kit (Affymetrix) and sequenced in both directions with the same primers used in the amplification step. Sequencing reactions were prepared with the Big Dye Terminator Cycle Sequencing Kit v1.1 (Applied Biosystems) and electrophoresed in an ABI 3730 DNA Analyzer (Applied Biosystems). The resultant chromatograms were edited, and all sequences were submitted to GenBank (GenBank accession numbers KM597073-KM597153).

A final dataset comprising the 81 Ugandan pig mitochondrial sequences obtained by us (Table S1) plus 183 sequences retrieved from GenBank (Table S2) was subjected to genetic analyses. Because not all of these 264 sequences were exactly the same length, they were trimmed, and an alignment of 464 bp was used to build a median-joining network with NETWORK 4.6 (Bandelt et al. 1999). We employed the default parameters implemented in NETWORK 4.6, and the transversion/transition ratio was changed to 3:1. This analysis (Fig. 1, Fig. S1) evidenced that East African pigs have a complex ancestry, with roots in Asia and the West (Europe/North Africa). However, the most original finding of our analysis was that Kenyan, Ugandan and Indian pigs cluster together (Fig. 1, Fig. S1). This result suggests a direct introgression event, because modern domestic pigs in India and Bhutan carry unique and highly differentiated mitochondrial haplotypes that can be found only in Indian wild boars (Larson *et al.* 2011). This finding is supported by previous data obtained in East African cattle, where zebuine alleles reach very high frequencies (Hanotte *et al.* 2002; Decker *et al.* 2014), probably because of maritime introductions associated with the Indian Ocean trade. Similarly, an Indian origin has been postulated for chicken in Zimbabwe on the basis of mitochondrial data (Muchadeyi *et al.* 2008; Mtileni *et al.* 2011).

The majority of Ugandan pigs grouped with the Far Eastern ones, but some of them fell within the European cluster (Fig. 1, Fig. S1). An Iberian influence on East African pigs is conceivable because the Portuguese were the first Europeans to circumnavigate Africa and establish a string of trading posts and colonies (Kilwa, Sofala, Mombasa, etc.) on its eastern coast (Okello 2002). Besides, standard European breeds, such as Large White and Landrace, also are widespread in the African continent because of their excellent productive abilities, which often overcome those of local populations. Clustering of Ugandan and Kenyan pigs with those from the Far East was also evident (Fig. 1, Fig. S1). At present, we do not know the source of this introgression because East African pigs clustered indistinctly with specimens from China and South East Asia. The presence of Indian and Far Eastern haplotypes in East African pigs might have been facilitated by the existence of trading routes (e.g., Carreira da Índia, Spice Trade, etc.) used by the Portuguese ships that, during the 15th to 18th centuries, connected China (Macau), Malaysia (Malacca) and India (Goa and Calicut) with Kenya (i.e., Malindi and Mombasa), Mozambique and other coastal regions of East Africa (Guinote 1999). Alternatively, this introduction may have taken place much before the Age of Discoveries, given



© 2015 Stichting International Foundation for Animal Genetics, 46, 433–436



**Figure 2** Neighbor-joining tree built with MEGA 5.05 (Tamura-Nei model, 1000 bootstrap replicates) of Y-chromosome *UTY* sequences from Ugandan pigs (blue) sampled at Masaka, Bundibujjo, Luweero, Rakai, Mpigi, Lira, Gayaza and Soroti. It can be seen that they all cluster with the *HY1* lineage (black), whereas the *HY3* (red) lineage appears as an outgroup.

that Indian Ocean trading routes have been used from immemorial times by the Arabs, Chinese, Persians, Romans, and many other civilizations (Kearney 2004).

Paternal genetic data also were obtained from Ugandan pigs by sequencing 0.37 kb of intron 1 and part of the flanking exons 1 and 2 of the ubiquitously transcribed tetratricopeptide repeat containing, Y-linked gene (UTY) with a single primer pair (forward 5'-AGCTGTTTTCGGTGAT GAGG-3', reverse 5'-TGCCCAACAGAGTTTTAGTCC-3'). Amplification reactions were carried out in a 15-µl final volume containing PCR buffer, 2.5 mM of MgCl<sub>2</sub>, 0.25 mM dNTPs, 0.3 µm of each primer and 0.75 of U Tag Gold DNA Polymerase (Applied Biosystems). The thermocycling profile consisted of one cycle at 95 °C for 10 min, followed by 35 cycles of 95 °C for 1 min, 60 °C for 1 min, 72 °C for 1 min and a final extension step at 72 °C for 7 min. The PCR products were purified with the ExoSAP-IT PCR Cleanup kit (Affymetrix) and sequenced following the same protocols employed for the mitochondrial control region. The UTY amplicon contains two diagnostic SNPs that allow differentiation of the two main Y-chromosome lineages, that is HY1 + HY2 vs. HY3 (Ramírez et al. 2009). After sequencing 18 Ugandan sires, we found only a single HY1 haplotype (Fig. 2) that is widely spread both in Asia and Europe. Contrary to our expectations, we did not find any trace of the HY3 haplotype, that could be considered as Asia specific and segregates in Kenyan (HY3 frequency: 35%) and Zimbabwean Mukota (HY3 frequency: 100%) pigs at significant frequencies (Ramírez et al. 2009). Probably, the impact of the Far Eastern introgression in African pigs has been especially important in Zimbabwe, where Mukota swine displaying a phenotype that resembles that of Chinese lard pigs have been reported (Ellert 1993). This genetic influence, in contrast, would have been less intense in other countries such as Uganda and Kenya.

In summary, this study demonstrates that pigs from East Africa have a very complex ancestry, with roots in the West, India and the Far East. This result is consistent with the enormous importance of East Africa as a point of intersection of intercontinental maritime routes linking these three geographic areas, a circumstance that has probably played a fundamental role in shaping and expanding African livestock diversity.

#### Acknowledgements

We would like to acknowledge the District Veterinary Officers working in the areas where samples were collected and the Ministry of Agriculture Animal Industry and Fisheries in Uganda for granting permission to carry out the study. We are greatly indebted to the Swedish International Development Agency (Sida) under the framework of Sida-Mak Bilateral Research Support Programme Phase 3 to the Post Doc program at the Directorate of Graduate and Research Training for the financial support (Grant No. 75007369, Swedish Research Links, contract number 348-2011-7380). Many thanks also to Karl Ståhl, for his collaboration in the project.

# References

- Amills M., Ramírez O., Galman-Omitogun O. & Clop A. (2013) Domestic pigs in Africa. African Archaeological Review 30, 73–82.
- Bandelt H.J., Forster P. & Röhl A. (1999) Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* **16**, 37–48.
- Blench R.M. (2000) A history of pigs in Africa. In: Origins and Development of African Livestock: Archaeology, Genetics, Linguistics and Ethnography (Ed. by R.M. Blench & K. Mac Donald), pp. 355– 367, Routledge Books, London, UK.
- Decker J.E., McKay S.D., Rolf M.M. *et al.* (2014) Worldwide patterns of ancestry, divergence, and admixture in domesticated cattle. *PLoS Genetics* **10**, e1004254.
- Ellert H. (1993) *Rivers of Gold (Zambeziana)*. Mambo Press, Gweru, Zimbabwe.
- Guinote P.J.A. (1999) Ascensão e Declínio da Carreira da Índia, Vasco da Gama e a Índia, vol II. pp 7–39, Fundação Calouste Gulbenkian, Lisboa.
- Hanotte O., Bradley D.G., Ochieng J.W., Verjee Y., Hill E.W. & Rege J.E. (2002) African pastoralism: genetic imprints of origins and migrations. *Science* 296, 336–9.
- Kearney M. (2004) The Indian Ocean in World History (Themes in World History). Routledge, London, UK.
- Larson G., Cucchi T. & Dobney K. (2011) Genetic aspects of pig domestication. In: *The Genetics of the Pig* (Ed. by M.F. Rothschild & A. Ruvinsky), pp. 14–37, CABI Publishing, Oxfordshire.

- Mtileni B.J., Muchadeyi F.C., Maiwashe A., Chimonyo M., Groeneveld E., Weigend S. & Dzama K. (2011) Diversity and origin of South African chickens. *Poultry Science* **90**, 2189–94.
- Muchadeyi F.C., Eding H., Simianer H., Wollny C.B., Groeneveld E. & Weigend S. (2008) Mitochondrial DNA D-loop sequences suggest a Southeast Asian and Indian origin of Zimbabwean village chickens. *Animal Genetics* 39, 615–22.
- Okello B. (2002) A History of East Africa. Fountain Publishers, Kampala, Uganda.
- Ramírez O., Ojeda A., Tomàs A. et al. (2009) Integrating Ychromosome, mitochondrial, and autosomal data to analyze the origin of pig breeds. *Molecular Biology and Evolution* 26, 2061–72.

### Supporting information

Additional supporting information may be found in the online version of this article.

**Figure S1** A more detailed view of the geographic origin of the samples included in the median-joining network shown in Fig. 1.

Table S1 Ugandan samples analyzed in the current studyand GenBank accession numbers of the correspondingmitochondrial control region sequences.

 Table S2 European and Asian mitochondrial control region sequences.